

## S32-1 Behavior of membrane receptors in living cells visualized by using a new tag-probe fluorescence labeling method

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To investigate behavior of membrane receptors in cell membranes, detection methods in living cells are essential. The specific labeling of proteins in living cells using a genetically encodable tag and a small synthetic probe targeted to the tag has several advantages over widely used fluorescent proteins. We recently developed a method using a high-affinity coiled-coil formation between the E3 tag (EIAALEK)<sub>3</sub> attached to the target protein and the Kn probes (KIAALKE)<sub>n</sub> (n = 3 or 4) labeled with a fluorophore [1]. The coiled-coil method is suitable for membrane proteins and having advantages such as 1) smaller size (1/5 of fluorescent proteins), 2) no impairment of receptor function, 3) easy control of labeling ratio, 4) cell-surface specific labeling 5) high specificity (K<sub>d</sub> = 6 nM and 64 nM for K4 and K3, respectively), 6) quick labeling within 1 min. The agonist-induced internalization of  $\beta$ 2-adrenergic receptor ( $\beta$ 2AR) and EGF receptor could be clearly visualized. Following internalization, pH of endosomes is known to decrease to 5–6. Fluorescence ratiometric detection of  $\beta$ 2AR internalization was examined. The detection simplicity compared with the conventional granularity analysis is useful for high throughput screening. [1] Yano, Y., Yano, A., Oishi, S., Sugimoto, Y., Tsujimoto, G., Fujii, N., Matsuzaki, K. (2008) ACS Chem. Biol. 3, 341–345.